

Guanidine Is a Zn²⁺-Binding Ligand at Neutral pH in Aqueous SolutionShin Aoki,[†] Kenta Iwaida,[†] Noriko Hanamoto,[†] Motoo Shiro,[§] and Eiichi Kimura^{*†}

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Arginine (Arg) residues having p*K*_a of 12.5 in proteins are perceived to function as guanidinium cations to bind to anions such as phosphates. Although acidic transition metals such as Zn²⁺ are common in metalloproteins and in metal-activated enzymes, their coordinate binding to Arg has not been proposed. Indeed, metal–guanidyl complexes are barely known and unexplored.^{1–3} A recent success of Pt²⁺–guanidyl complexes at pH 7.5 was ascribed to specific circumstances; that is the highly acidic nature of Pt²⁺ lowered p*K*_a values of guanidiniums by hydrophobic environment or π acceptor (guanidine)–π donor (Pt²⁺) interaction.^{2,3} Herein we report that guanidine can be a good ligand to Zn²⁺ in neutral pH aqueous solution.

A new ligand, (2-guanidinyl)ethyl-cyclen (L¹) **1** formed a stable 1:1 Zn²⁺ complex (ZnL¹) **2b**, which crystallized out as a 2ClO₄[−] salt from pH 7.5 aqueous solution (Scheme 1).⁴ X-ray crystal structure analysis revealed a distinct coordination of the pendant guanidine (through N18) to Zn²⁺ (Figure 1). Zinc(II) is thus five-coordinate with four nitrogens of the cyclen ring (the average Zn²⁺–N bond distance is 2.16 Å) and a nitrogen of guanidine (1.95 Å). A shorter bond length (1.31 Å) between C17 and N18 with respect to those for C17–N16 (1.35 Å) and C17–N19 (1.35 Å) implies that the Zn²⁺-bound N18 is an imine nitrogen.¹

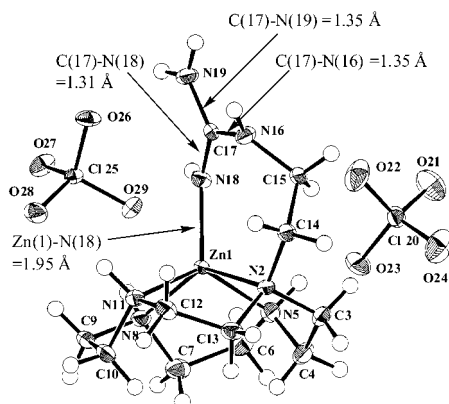


Figure 1. ORTEP drawing (50% probability ellipsoids) of ZnL¹ complex (**2b**). Selected bond distances (Å): Zn1–N2 2.188(2), Zn1–N5 2.125(2), Zn1–N8 2.180(1), Zn1–N11 2.135(2), Zn1–N18 1.953(2), C17–N16 1.352(2), C17–N18 1.310(2), C17–N19 1.347(2).

How does the guanidine in **2** prefer Zn²⁺ over protons at neutral pH? From potentiometric pH titration of the ligand L¹ (1 mM) in the absence and presence of 1 mM Zn²⁺ at 25 °C with *I* = 0.1 (NaNO₃), the p*K*_a values of >12, 10.13, 8.45, <2, and <2 were determined by the program “BEST”.^{5,6} In light of the p*K*_a values of 10.7, 9.7, <2, and <2 for cyclen (L²),⁷ the highest p*K*_a value of

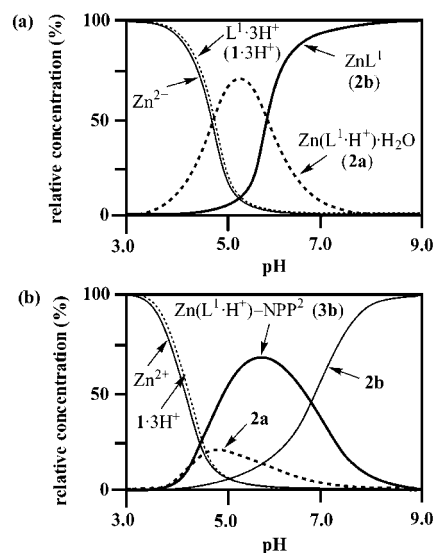


Figure 2. (a) Speciation diagrams for a mixture of 1 mM **1** + 1 mM Zn²⁺ as a function of pH at 25 °C with *I* = 0.1 (NaNO₃). (b) Speciation diagrams for a mixture of 1 mM **2** + 1 mM NPP as a function of pH at 25 °C with *I* = 0.1 (NaNO₃). Other species in less than 5% are omitted for clarity.

> 12 for L¹ was assigned to the pendant guanidinium ion. In Scheme 1, the Zn²⁺ complexation constant (log *K* (Zn(L¹·H⁺)) for **1**·H⁺ + Zn²⁺ ⇌ Zn(L¹·H⁺)·(H₂O) (**2a**) and the deprotonation constant p*K*_a (Zn(L¹·H⁺)) for **2a** ⇌ **2b** + H₃O⁺ were 12.4 ± 0.1 and 5.9 ± 0.1, respectively, at 25 °C with *I* = 0.1 (NaNO₃).⁸ Zn²⁺-cyclen complexes are good models of active centers of various zinc(II) enzymes.⁹ For instance, the deprotonations from a Zn²⁺-bound water (**4a** ⇌ **4b**)¹⁰ and from a Zn²⁺-bound alcohol (**5a** ⇌ **5b**)¹¹ are facile with the p*K*_a values of 7.9 and 7.7, respectively. It is thus reasonable that the deprotonation of the guanidinium for **2a** ⇌ **2b** is facile due to the close interaction between the Zn²⁺-bound water (or OH[−]) and the guanidinium ion in **2a**. An apparent complex formation constant, log *K*_{app}(ZnL¹) was calculated to be 10.4 in comparison to 11.0 for ZnL² and 10.4 for ZnL³ at pH 7.4.¹² The speciation diagram for a mixture of 1 mM L¹ and 1 mM Zn²⁺ as a function of pH at 25 °C with *I* = 0.1 (NaNO₃) indicates that the initially formed Zn²⁺ complex Zn(L¹·H⁺)·H₂O **2a** is populated most abundantly (75%) at ca. pH 5.5 (Figure 2a).

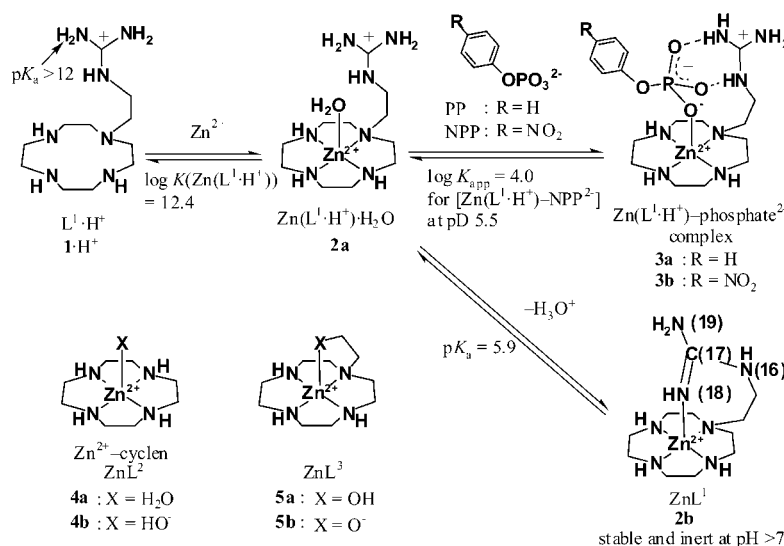
Another interesting question with the Zn²⁺–guanidine bonding in **2b** was how labile is it in aqueous solution? We found that **2b** did not catalyze the hydrolysis of 4-nitrophenyl acetate at pH 5.0–9.0 (see the Supporting Information), unlike the catalytically reactive nucleophiles **4b**¹⁰ and **5b**.¹¹ We then tested if an external (4-nitrophenyl)phosphate (NPP) anion could displace the pendant guanidine in **2b**. The dianionic NPP was a good ligand to Zn²⁺ in **4** to form an 1:1 complex with log *K*_{app} = 3.1 at pH 5.6.¹³ Figure 3 compares the ³¹P NMR (162 MHz) titration curves of NPP (5

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Scheme 1



mM) with **1**, **2**, and **4** at various pD, from which a 1:1 complex **3b** formation with $\log K_{app}$ (**2**-NPP) of 3.7 ± 0.1 at pD 5.5 ± 0.1 was calculated.¹⁴ A structure of **3b** was assigned, as depicted in Scheme 1. The ligand **1** alone (mostly in the $L^1 \cdot 2H^+$ form) had little interaction with NPP at pD 5.5 ± 0.1 , see Figure 3.¹⁵ On the basis of potentiometric pH titration, a speciation diagram was obtained for a mixture of 1 mM **2** and 1 mM NPP as a function of pH at 25 °C with $I = 0.1$ (NaNO₃) (Figure 2b).⁶ At higher pH, the pendant guanidinium became deprotonated to displace the Zn^{2+} -bound NPP²⁻ to yield **2b**. The apparent complexation constant, $\log K_{app}$ (**2**-NPP), of 4.0 ± 0.1 at pH 5.5 agreed reasonably well with 3.7 ± 0.1 obtained by the ³¹P NMR method. A higher affinity of NPP²⁻ to Zn^{2+} in **2a** over **4a** probably arose from the phosphate-guanidinium interactions, as depicted in **3b**.

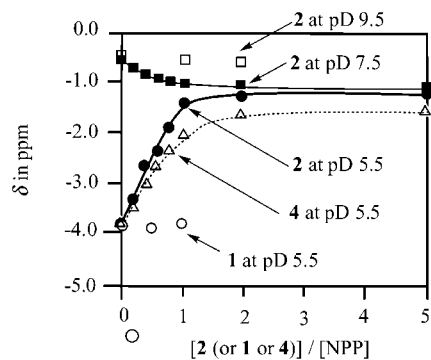


Figure 3. pH-dependent change (at pD 5.5, 7.5, and 9.5) of ³¹P chemical shift of NPP (5 mM) upon addition of **1**, **2**, and **4** in D₂O at 35 °C.

Colorless prisms were obtained from a mixture of **2b** and phenyl phosphate (PP) in aqueous solution at pH 6.5. The X-ray crystal structure analysis proved the 1:1 **2**-PP²⁻ complex (**3a**), where the Zn^{2+} -bound PP²⁻ is stabilized by hydrogen bondings with two intermolecularly adjacent guanidinium protons (see the Supporting Information). The intermolecular interactions may come from strong crystal packing forces. The present facts may imply another function of Arg residues in the active centers of zinc(II) enzymes.^{16,17}

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Supporting Information Available: Experimental section, Figures S1–S6, and Tables and CIF for **2b** and **3a** (PDF). This material is available free of charge via the Internet at <http://pubs.acs.org>.

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 $K(Zn(L^1 \cdot H^+)) = [Zn(L^1 \cdot H^+) \cdot H_2O] / [Zn^{2+}][L^1 \cdot H^+] \text{ (M}^{-1}\text{)}$ (1)
 $2a \rightleftharpoons 2b + H^+$: $K_a(Zn(L^1 \cdot H^+)) = [2b]a_{H^+} / [2a]$ (2)
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- (12) The $K_{app}(ZnL^m)$ values are defined by eqs 3 and 4. For detailed comparison, see the Supporting Information.
 $K_{app}(ZnL^m) = [2a + 2b \text{ (or } 4a + 4b \text{ or } 5a + 5b)] / [Zn^{2+}]_{free} [L^m]_{free}$ (at designated pH) (M⁻¹) (3)
 $[L^m]_{free} = \sum [L^m \cdot nH^+]_{free}$ (n = 0–5 for m = 1 and 3, and n = 0–4 for m = 2) (4)
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- (14) The pK₁ and pK₂ values for NPP are <2 and 5.2 ± 0.1 , respectively, at 25 °C with $I = 0.1$ (NaNO₃) (ref 13a, b).
- (15) The ³¹P NMR titration for NPP (5 mM) and **4** was carried out at pD 5.5 ± 0.1 and 35 °C, giving $\log K_{app}$ (**4**-NPP) of 3.4 ± 0.1 (Figure 3).
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